

# Transportation Environment of Contaminated Instruments before Reprocessing

Karin Bundgaard, Associated Professor, PhD, MScN, RN Aalborg University Hospital & Aalborg University





### Partners in crime



Peter Rubak



Gerhard Kirmse



Silke Haibt-Winandi





### The context









# Introduction

### International infection control guidelines recommend

- Reprocessing should commence immediately after use
- Different recommendations for environment
  - Europe: mostly dry to minimize risk of corrosion
  - UK and America: mostly moist to ensure efficient cleaning outcomes

#### However

- Recommendations are based on best practice and expert knowledge
- Research in a real-life clinical setting is warranted





Can a humid storage environment of surgical instruments before reprocessing increase patient safety and durability of instruments?

P. Rubak  $^a,$  J. Lorenzen  $^b,$  K. Ripadal  $^c,$  A-E. Christensen  $^d,$  D. Aaen  $^e,$  H.L. Nielsen  $^{e,f},$  K. Bundgaard  $^{f,g_{e^*}}$ 





# Aim

### Aim of the study

To compare moist and dry transportation of surgical instruments in a real-life clinical setting

- Protein residues (cleanliness)
- Corrosion (surface changes)







### **Data collection**

- Aarhus University Hospital
- Basic instrument sets (54 and 39 surgical instruments)
- Transportation
  - Dry: Abdominal Surgery Department
  - Moist: Orthopedic Spine Surgery Department

### **Defining Moist transportation/storage**

- OR-Gauze (16 g, 30x45 cm) soaked with 300 ml of sterile water
- Closed container











#### **Protein residue test**

- 5-10 contaminated inst each tray
- Washing process as is ( the disinfection phase)
- Elution process (include
- High-sensitive BCA prot (Bicinchoninic Acid)

PF-12"" ()

al cleaning/disin	fection
ng	2 min
nzymatic Getinge cleaner : 3 ml/l)	5 minutes at 55 <sup>0</sup> C (total cleaning time 8 minutes)
	Soft water
	Soft water
lisinfection	Demineralized water A <sup>0</sup> 3000
	15 minutes







### Calibration curve BCA Protein Assay Kit









### Surface changes

- Each instrument was visually inspected
  - Contact corrosion
  - Fretting
  - Pitting corrosion
  - Stains
  - Residue

|--|

0	No corrosion
1	Single small corrosion spot
2	Larger single corrosion spot, pitting
3	Several small corrosion spots
4	Several larger corrosions spots
5	Massive corrosion, multiple spots







# Fretting corrosion



1 point

2 point







### **Contact corrosion**



1 point

2 point

3 point





# Pitting corrosion



1 point





2 point

3 point





# Residue







# Staining







## General data – Protein residue

#### **Reprocessing Cycles**

	MTK	, Dry	EOP, Moist		
	Total Tested		Total	Tested	
Trays	24 12		10	8	
Cycle, Mean	76	75	67	66	
Cycle, Min	58	58	55	55	
Cycle, Max	99	99	70	70	

### Waiting time before reprocessing

	Number of trays			
	Dry Moist			
2-4 hours	0	1		
4-8 hours	1	1		
8-12 hours	1	1		
12-24 hours	8	6		







### General data - Corrosion

#### **Reprocessing Cycles**

	MTK	, Dry	EOP, Moist		
	Total Tested		Total	Tested	
Trays	24 10		10	9	
Cycle, Mean	76	75	67	66	
Cycle, Min	58	58	55	55	
Cycle, Max	99	99	70	70	

#### **Exchanged instruments**

	Number	Trays
Dressing Forceps	2	2
Forceps (Clamp)	3	2
Needleholders	5	5
Retractors	4	3
Scissors	6	5
Tweezers	4	2
Total	24	







#### **Protein residue**



Numbers above the bars represents number of instruments

	Dry	Moist
n	89	84
Mean, [µg/Probe]	27,7	26,9
Median, [µg/Probe]	10,0	15,0
Min, [µg/Probe]	10	10
Max, [µg/Probe]	336	333
Sd	43,5	42,6
CV%	63,6	63,0
Nr. of trays	12	8

- Kruskal-Wallis test (raw data) p=0.56
- Chi<sup>2</sup>-test (categorized data) p=0.55





#### Surface changes



Numbers above the bars represents number of instuments





### **Corrosion grading including fretting**



	Dry	Moist
n	507	349
Mean, Points	0.8	1.3
Median, Points	1.0	1.0
Nr. of trays	10	9





### **Corrosion grading without fretting**



	Dry	Moist
n	507	349
Mean, Points	0.4	1.0
Median, Points	0.0	1.0
Nr. of trays	10	9





### **Pitting and Stains**

	Dry	Moist
n	507	349
Pitting	9	10
Stains	68	47
Nr. of trays	10	9







### Corrosion with and without fretting on scissors







### **Corrosion with and without fretting on forceps**







### **Comparison of instrument types**

	n		Corrosion w. fretting, %		Corrosion w.o. fretting, %	
	Dry	Moist	Dry	Moist	Dry	Moist
Forceps	217	126	50	80	27	45
Needleholders	46	35	57	83	13	66
Scissors	64	36	84	97	44	92
Tweezers	56	72			45	85





# Conclusion

### **Storage Environment and Instrument Cleanliness**

- No correlation between storage environment and level of protein residue
- Insufficient data to establish a link between instrument type and level of protein residue

#### **Storage Environment and Corrosion Formation**

- Moist storage environments results in higher corrosion

#### **Corrosion Across Instrument Types**

- Consistency across instrument types independent of storage environment
- Scissors are the most corroded compared to needleholders, forceps, and tweezers

### **Reprocessing cycles and surface changes**

- No correlation between number of reprocessing cycles and surface changes





# Strengths and Limitations

- Real life setting
- Instruments used for surgery
- Number of examined instruments
- Reprocessing: Standard protocols for washing, disinfection and sterilization
- Handling of reprocessing: Trained personnel from the CSSD
- Protein residue analysis: Performed by professionals and use of a high sensitivity BCA method
- Corrosion analysis: Performed by professionals and use of a standardized scoring system
- Choice of method of creating a moist environment spray instead of sterile water
- Choice of instruments complex instruments instead of basic instruments





# Thank you!